OCCURRENCE OF MYCOBIOTA IN EASTERN AUSTRALIAN SEA TURTLE NESTS

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Major loggerhead (Caretta caretta), green (Chelonia mydas), hawksbill (Eretmochelys imbricata) and flatback (Natator depressus) sea turtle rookeries in eastern Australia were surveyed for the presence of fungi in turtle nests. While the fungi Fusarium oxysporum, F. solani and Pseudallescheria boydii were not universally present, they were isolated from the exterior of failed eggs of all turtle species and at all rookeries. Predominant fungi species varied spatially and temporally. Sea turtle, egg, nest. fungi.

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Globally, fungi have been described from the exterior and/or embryonic tissue of several species of sea turtle eggs (Table 1). Fusarium oxysporum Schlecht., F. solani (Mart.) Sacc. and Pseudallescheria boydii Negroni & Fischer have been identified from the exterior of failed eggs in sea turtle nests at Heron Island (Phillott et al., 2001). There have been no studies to determine the identity of fungi from failed eggs on turtle rookeries along a single coastlinc.

Heron Island (23°26'S, 151°55'E), Wreck Island (23°21'S, 151°57'E), Peak Island (23°20.5'S, 150°56'E), Milman Island (11°10'S, 143°00'E) and Mon Repos Conservation Park (24°48'S. 152°27'E) are major rookeries for the loggerhead (Caretta caretta), green (Chelonia mydas), hawksbill (Eretmochelys imbricata) and flatback (Natator depressus) turtles. These were surveyed in the 1996/97-1998/99 turtle nesting seasons (Table 2). Nests were located after hatchling emergence and excavated by hand to determine hatch success. A single unhatched egg that appeared by visual inspection to be colonised by fungus was taken from some nests (Table 2) and swabbed using MW170 TRANSTUBE®, Amies Clear Media and refrigerated at 3-5°C. Fungal swabs were incubated on half-strength Potato Dextrose Agar with 0.05gL⁻¹ chloramphenicol to inhibit bacterial growth and then subcultured onto a range of media (e.g. Potato Dextrose Agar, Nutrient Agar, Carnation Leaf Agar) as required for identification. Identification of species not included by Booth (1971) and Seifert (1996) was confirmed by Professor D.E. Ellis. Voucher specimens were lodged with the Queensland Department of Primary Industries (Accession

Numbers: F. oxysporum BRIP 28368, F. solani BRIP 28369, P. boydii BRIP 28370).

F. oxysporum, F. solani and P. boydii were isolated from sea turtle nests at all rookeries investigated (Table 2). The proportion of nests containing each fungal species varied between rookeries and, at Heron I. (the only rookery to be sampled in multiple years), between nesting seasons. Most samples were of single isolates, however 6.54% of all swabs revcaled mixed cultures (1 F. oxysporum + F. solani; 2 F. oxysporum + P. boydii; 4 F. solani + P. boydii). While individual rookery/species census data often show absence of a particular fungus, all fungi were recorded from all turtle species.

As all 3 of the mycobiota isolated from Heron I. (Phillott et al., 2001) are common soil saprophytes, their presence in nests at other turtle rookerics in eastern Australia is not surprising. The temporal variation in predominant fungi observed at Heron I. (1996: F. solani; 1997: P. boydii) suggests that spatial differences in species composition cannot be solely attributed to geographic location, as it may equally have been influenced by environmental factors (e.g. hydric and thermal) and/or temporal variation.

A comparison of rainfall distribution and volume (1996: 237mm; 1997: 183mm), and air temperature (± SD 1996: max. 29.8±1.7°C, min. 23.1±1.6°C; 1997: max. 30.9±2.0°C, min. 24.0±1.9°C; beach temperature data not available), differed at Heron I. over the November to February sampling period between the 2 seasons. This may have contributed to the change in dominant nest mycobiota, as *F. solani* is more prevalent in areas with relatively high rainfall and low temperature (Burgess &

TABLE 1. Records of fungi associated with failed sea turtle eggs.

Summerell, 1992). The germination rate and hyphal growth of all 3 fungi at varying hydric and thermal microclimatic conditions (experienced within natural sea turtle nests) is under further investigation.

Loggerhead turtle nesting populations in Queensland are in rapid decline (50-80% over 10-15yrs dependent on study site: Limpus & Reimer, 1994). Loggerhead nesting in the south Pacific Ocean occurs almost entirely in the southern Great Barrier Reef (the most significant being at Wreck I.) and Bundaberg coastline (including Mon Repos Conservation Park: Limpus, 1985). However loggerhead nests demonstrate high mortality (and associated fungal presence) on Wreck and Heron Islands when compared with green turtle nests at the same location, or loggerhead turtle nests on the adjacent mainland (Limpus et al., 1983; Phillott, 2002). Determining the role of fungi as an opportunistic contaminant of failed turtle eggs or infectant of living embryonic tissue is, therefore, of great conservation importance.

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LITERATURE CITED

ACUÑA-MESÉN, R.A. 1992. Monosporium apiospermum Saccardo (Fungi, Deuteromycetes), asociado a los hucvos de la tortuga marina Lepidochelys olivacea (Eschscholtz 1829) en Costa Rica. Brenesia 38: 159-162.

BOOTH, C. 1971. The genus *Fusarium*. (Commonwealth Mycological Institute: Kew).

BURGESS, L.W. & SUMMERELL, B.A. 1992. Mycogeography of *Fusarium*: survey of *Fusarium* species in subtropical and semi-arid grassland soils from Queensland, Australia. Mycological Research 96: 780-784.

ECKERT, K.L. & ECKERT, S.A. 1990. Embryo mortality and hatch success in *in situ* and translocated leatherback sea turtle *Dermochelys coriacea* eggs. Biological Conservation 53: 37-46

LIMPUS, C.J. 1985. A study of the Loggerhead Turtle, *Caretta caretta*, in Eastern Australia. Unpubl. PhD thesis, University of Queensland, St Lucia.

LIMPUS, C.J. & REIMER, D. 1994. The loggerhead turtle, Caretta caretta, in Queensland: a population in decline. Pp. 34-54. In James, R. (ed.) Proceedings of the Australian Marine Turtle Conservation Workshop, November 1990. (Australian Nature Conservation Agency: Canberra).

LIMPUS, C.J., REED, P. & MILLER, J.D. 1983. Islands and turtles. The influence of choice of nesting beach on sex ratio. Pp. 397-402. In Baker, J.T., Sammarco, P.W. & Stark, K.P. (eds) Proceedings of the Inaugural Great Barrier Reef Conference. (JCU Press: Townsville).

MO, C.L., SALAS, I. & CABALLERO, M. 1990. Are fungi and bacteria responsible for olive ridley's egg loss? Pp. 249-252. In Richardson, T.H., Richardson, J.I. & Donnelly, M. (comps) Proceedings of the 10th Annual Workshop on Sea Turtle Biology and Conservation. NOAA Technical Memorandum NMFS-SEFC-278.

PETERS, A., VERHOEVEN, K.J.F. & STRIJBOSCH, H. 1994. Hatching and emergence in the Turkish Mediterranean loggerhead turtle, *Caretta caretta*: a natural cause for egg and hatchling failure. Herpetologica 50: 369-373.

TABLE 2. Spatial and temporal variation in fungal presence in sea turtle nests.

Rookery, Season	Turtle	# Nests Excavated	# Nests Swabbed	% Nests		
				F. oxysporum	F. solani	P. boydii
Heron I. 1996/97 Heron I. 1997/98	Green Loggerhead Green Loggerhead	86 2 108 5	22 2 19 3	22.7 0.0 5.0 0.0	63.6 100.0 5.0 0.0	13.6 0.0 100.0 100.0
Milman 1. 1998/99	Green Hawksbill	7 32	5 8	40.0 12.5	60.0 75.0	0.0 12.5
Mon Repos 1998/99	Loggerhead	12	5	20.0	100.0	20.0
Peak 1. 1998/99	Flatback	44	16	12,5	62,5	25.0
Wreck I. 1998/99	Green Loggerhead	39 7	16 4	18.8 50.0	62.5 50.0	37.5 25.0

PHILLOTT, A.D. 2002. Fungal colonisation of sea turtle nests in Eastern Australia. Unpubl. PhD thesis, Central Queensland University, Rockhampton, Queensland.

PHILLOTT, A.D., PARMENTER, C.J. & LIMPUS, C.J. 2001. Mycoflora identified from failed green (*Chelonia mydas*) and loggerhead (*Caretta caretta*) sea turtle eggs at Heron Island, Australia. Chelonian Conservation and Biology 4: 170-172.

SEIFERT, K. 1996. FusKey: Fusarium Interactive Key. http://www.res.agr.ca/brd/fusarium.

SOLOMON, S.E. & BAIRD, T. 1980. The effect of fungal penetration on the eggshell of the green turtle. Pp. 434-435. In Brederoo, P. & de Priester, W. (eds) Proceedings of the Seventh European Congress on Electron Microscopy. (7th European Congress on Electron Microscopy Foundation: Leiden).

WYNEKEN, J., BURKE, T.J., SALMON, M. & PEDERSON, D.K. 1988. Egg failure in natural and relocated sea turtle nests. Journal of Herpetology 22: 88-96.